



# A novel natural surface-enhanced Raman spectroscopy (SERS) substrate based on graphene oxide-Ag nanoparticles-Mytilus coruscus hybrid system

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## ABSTRACT

Recently, biological materials have attracted more and more attentions on preparing the surface-enhanced Raman spectroscopy (SERS) substrate with the conspicuous SERS performance. Herein, we demonstrate a simply facile and inexpensive approach to fabricate a novel natural SERS substrate based on graphene oxide-Ag nanoparticles-Mytilus coruscus (GO-AgNPs-M.c.). The surface-enhanced resonance Raman spectroscopy (SERRS) signals of the Rhodamine 6G (R6G) on the GO-AgNPs-M.c. hybrids exhibit the much better SERRS performances in terms of the sensitivity, the signal reproducibility and the antioxidant stability than that on the AgNPs-M.c. one. The well electric field enhancement contribution is likewise confirmed in theory via the COMSOL software. Moreover, the successful detection of trace malachite green (MG) and methylene blue (MB) indicates the well feasibility for multi-molecules detection on the GO-AgNPs-M.c. hybrids. Thus, this inexpensive and high-efficiency natural SERS substrate have great potential for the considerable biochemical SERS applications and broaden the way for preparing multiple SERS platforms derived from other natural materials.

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## 1. Introduction

Surface enhanced Raman spectroscopy (SERS), as a versatile analytical technique which could provide single molecular sensitivity, has attracted increasing attentions of both theoretical and experimental researchers in recent decades [1–4]. Due to the ultra-sensitive and label-free detection of chemical and biological molecules, SERS has shown great potential for practical applications in diverse fields, such as environmental monitoring, food safety, biomedical diagnostics, gas sensors and explosives in military [5–10]. It is generally accepted that electromagnetic mechanism (EM) and chemical mechanism (CM) are the major mechanisms of SERS enhancement [7,11]. The EM, originating from the dramatic increase of local electromagnetic field in the plas-

monic “hot spots” supported by strong coupling of localized surface plasmon resonance (LSPR), can provide a far higher enhancement factor than the CM which results from the charge transfer between the probing molecules and the substrate [12–14]. It is commonly believed that the noble metal with hybrid plasmonic structures can offer a prominent SERS effect from the EM. Thence, the efficient plasmonic nanostructures or configurations have been designed to achieve the highly sensitive SERS detection of trace molecules [11,15]. Diverse technologies (such as lithography techniques, nanosphere masks, self-assembly, porous silicon template etc.) have been emerging for developing the SERS substrate with the homogeneous plasmonic arrays [16–21]. However, the facile and inexpensive preparation of the meaningful SERS substrate with uniformly well-ordered arrays remains to be a significant challenge.

Recently, systematic studies have confirmed that certain biological materials possess the unique structural properties which could be applied in the material science [22,23]. In terms of the fascinating micro/nanostructures and the low-cost acquirement of the biological materials, the novel natural SERS substrates have been emerging endlessly [24–28]. Considerable research efforts have been investi-

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gated that these biological templates simply decorated with the plasmonic metals could show the conspicuous SERS effect for multi-molecules detection. For example, Mu and co-workers in situ synthesized gold nanoparticles on the surface of several butterfly wings with natural photonic architectures, achieving the detection of  $10^{-9}$  M 4-aminothiophenol (4-ATP) [24]. According to Shao's work, the novel platform with hierarchical nanogaps was designed by performing the cicada wings as bioscaffold arrays and successfully applied in the animal virus biosensing [26]. Besides, Huang and colleagues fabricated the natural substrate based on silver deposited taro leaf, which obtained highly sensitive and well reproducible signals by means of natural superhydrophobicity and rich hot spots provided by crossed nanoplate [27]. While these works achieve the well SERS performance based on the natural arrays, several disadvantages remain to exist. First, the poor manipulity of the fabricated natural SERS substrate greatly limits the practical applications. Since these biological materials are fragile, easily damaged during the fabrication process and the experimental measurement. Second, seasonal restriction and biological devastation of natural materials have prevented substrates from being prepared promptly to certain extent. Third, the oxidation of the noble metal (especially silver) directly decreases the long-term endurance of natural SERS substrates. Therefore, the more suitable natural template still need to be researched for fabricating the SERS substrate achieving the highly sensitive measurements with the excellent reproducibility and the well stability.

*Mytilus coruscus* (M.c.), belonging to mytilidae, is an extremely common edible aquatic mollusk. The shell of M.c. has been mostly thrown away after eating and become almost useless. Yet, recent research efforts have suggested that the internal surface of the shell exhibits the extremely exciting microstructures and may be divided into three regions: nacre region (N) region, adductor muscle scar (AMS) region and fibrous prism (FP) region [29–31]. More surprisingly, these unique microstructures on the M.c. are most likely to be utilized to prepare the plasmonic structures for the SERS measurement, including several advantages: excellent uniform microstructures in large scale, the high manipulity due to the antifrangible property, easily massive obtainment at low cost.

Graphene, as a typical two-dimensional nanomaterial with superior optical and electronic properties, has been increasingly applied to the SERS substrate, since it can further enlarge the SERS signals on account of the CM and suppress the fluorescence background from the probing molecules [32–35]. Compared to graphene, graphene oxide (GO) has rich oxygen-containing groups, which are more critical for the selective adsorption of molecules and the achievement of the SERS signals with well homogeneity, chemical stability and bio-compatibility [36–38]. Moreover, the GO can be used as the additional protective layer to maintain the stability of the SERS substrate [39].

In this paper, by combining the above advantages, we reported a novel natural SERS substrate based on graphene oxide-Ag nanoparticles-*Mytilus coruscus* (GO-AgNPs-M.c.). With R6G as the analyte molecules, the SERRS behavior of the three regions (N, AMS and FP) was firstly analysed on the non-decorated AgNPs-M.c. substrate. The FP region exhibited the best SERRS behavior due to abundant hot spots around the three-dimension (3D) active plasmonic natural structures. Furthermore, the GO-AgNPs-M.c. hybrids were fabricated by dip-coating the GO film on the AgNPs-M.c. hybrids, which achieved the more sensitive, reproducible and stable SERRS signals compared with the non-decorated AgNPs-M.c. hybrids. The minimum detectable concentration for R6G on the GO-AgNPs-M.c. substrate can reach  $10^{-14}$  M. The well electric field enhancement distribution is further confirmed by the COMSOL software. Experimental results demonstrate that the well-ordered natural arrays and the homogeneous decoration of GO lead to the excellent reproducibility and the well antioxidant stability of SERRS

signals. What's more, the successful detection of malachite green (MG) and methylene blue (MB) indicates that the GO-AgNPs-M.c. hybrids show the immense potential for practical applications in diverse biochemical fields.

## 2. Experimental

### 2.1. Preparation of the AgNPs-M.c. and GO-AgNPs-M.c. substrates

Fig. 1 schematically illustrates the fabrication processes of the AgNPs-M.c. and GO-AgNPs-M.c. substrates. The M.c. was collected from Rizhao, located at the seaside of Huanghai Sea in China. The length of selected shell for the experiment is approximately 6 cm. After the remove of the whole soft tissue by a knife, the shell was firstly treated with the alkali dissolution method, i.e. immersing the shell into the NaOH aqueous solution (20%, w/v) under the water bath  $65^{\circ}\text{C}$  for 1 h to remove the organic contaminants adhered to M.c. and eliminate the cuticle and partial organic membrane. The shell was subsequently put into deionized water bath sonication for 10 min twice, then dried in the  $\text{N}_2$  atmosphere. Afterwards, 0.001 g silver wire was deposited on the M.c. by the physical vapor deposition (PVD) method, forming the AgNPs-M.c. substrate. Furthermore, the  $20\ \mu\text{L}$  GO suspension with the concentration of 0.2 mg/mL was dip-coated on the AgNPs-M.c. substrate, fabricating the GO-AgNPs-M.c. substrate. The prepared AgNPs-M.c. and GO-AgNPs-M.c. substrates were sealed in the vacuum environment for the following SERS measurements. What needs illustration is that we merely deposited AgNPs at the whole M.c. surface in order to compare the surface morphology and the SERS performance of three regions. With respect to all other measurements, only FP region was separated from the M.c. by a knife to fabricate the AgNPs-M.c. and GO-AgNPs-M.c. substrates.

### 2.2. SERS experiments

The R6G solution obtained by dissolving the R6G in the water was used to test and compare the SERRS performances of different regions on the AgNPs-M.c. substrate and different prepared substrates ( $\text{SiO}_2$ , AgNPs-M.c. and GO-AgNPs-M.c.). The malachite green (MG) and methylene blue (MB) from  $10^{-5}$  to  $10^{-9}$  M were also used to detect the SERS effect of GO-AgNPs-M.c. substrate.  $2\ \mu\text{L}$  of these probe molecules solution was dropped directly on the tested substrates. Then, these samples were totally dried before performing the SERS experiments. The mentioned SERS spectra were collected with a Horiba HR Evolution 800 Raman spectrometer system (laser wavelength at 532 nm, 4.8 mW laser power, gratings with 600 grooves per 1 mm,  $\times 50$  objective lens,  $1\ \mu\text{m}$  laser spot, 8 s acquisition time etc.). These SERS spectra are completely the average of spectra randomly collected from at least 8 points on all substrates.

### 2.3. Characterization

The surface structures of all the substrates were characterized by scanning electron microscope (SEM, Zeiss Gemini Ultra-55). The energy dispersive spectroscopy (EDS) was utilized for the chemical elemental analysis. All Raman spectra were carried out with a Raman spectrometer (Horiba HR Evolution 800).

## 3. Results and discussion

As shown in Fig. 1, the image of M.c. can be seen clearly, the blue circle from left to right represents the N, AMS and FP regions, respectively. The surface morphologies of three regions were further investigated by the SEM images. As shown in Fig. 2(a) and (b), the morphology and cross-sectional SEM images of the N region

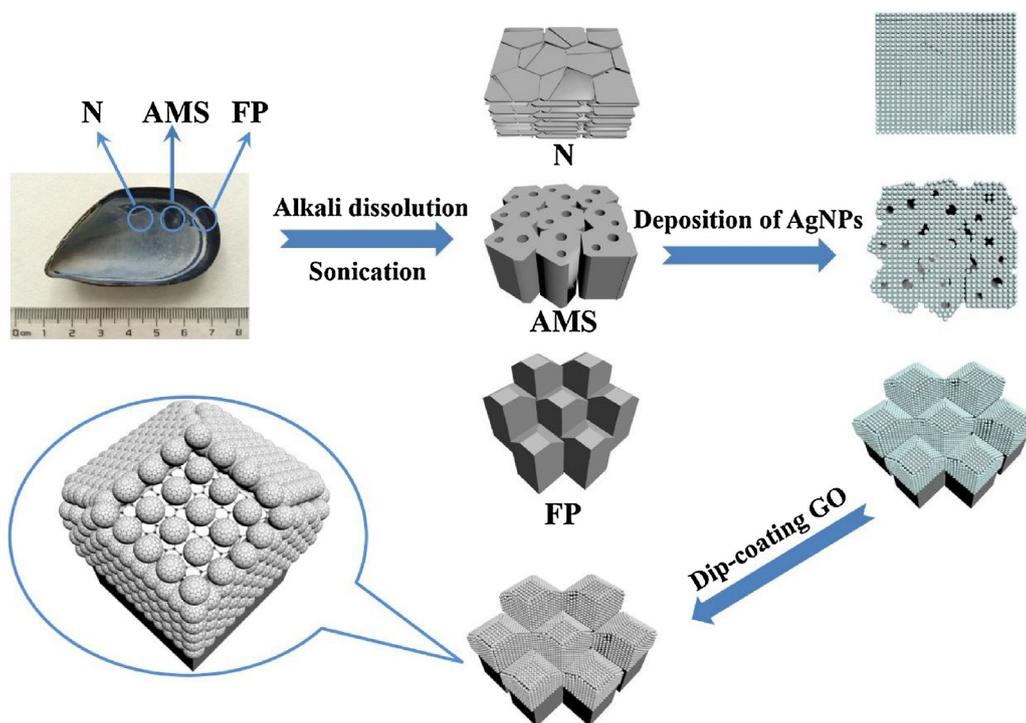


Fig. 1. Preparation processes of the AgNPs-M.c. and GO-AgNPs-M.c. substrates.

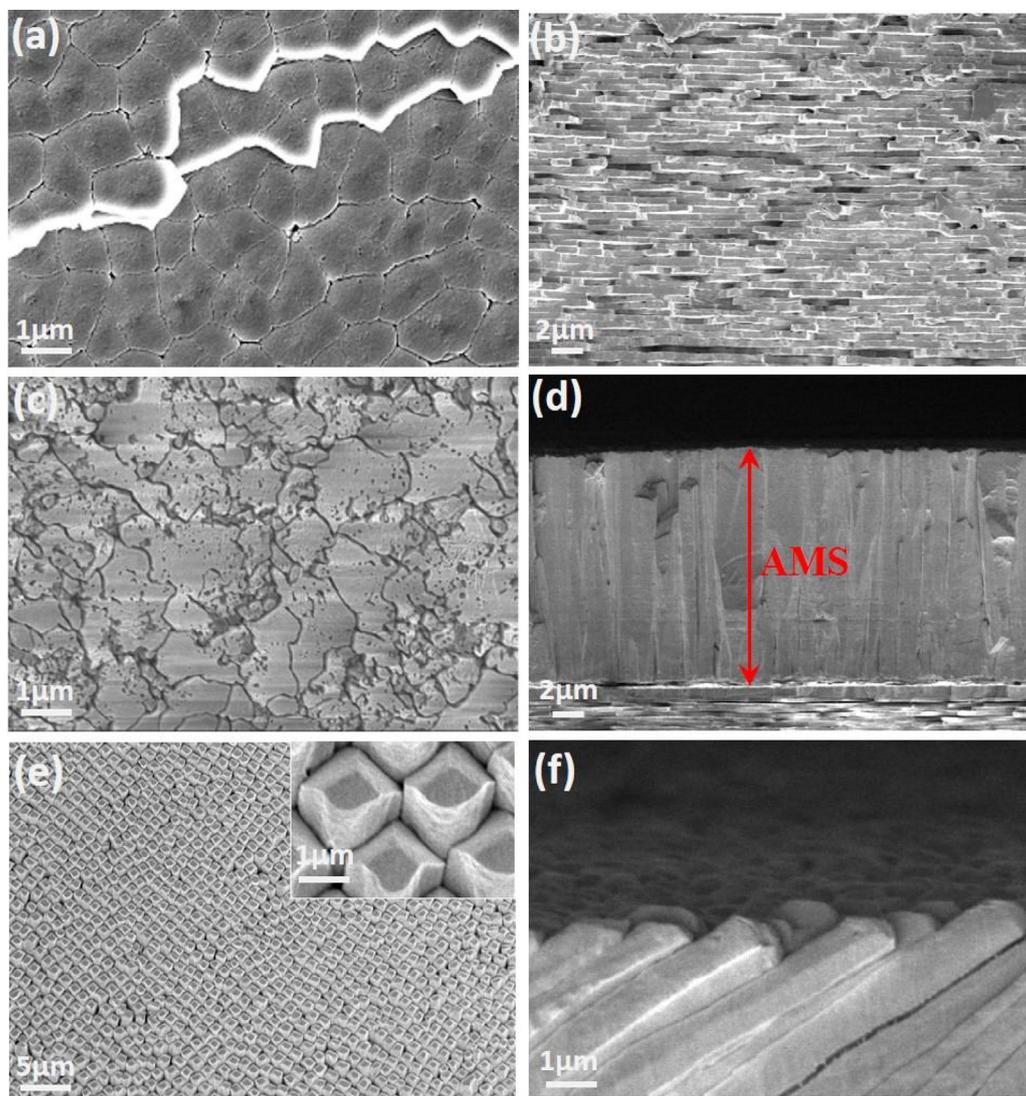
prove that the N region consists of the neat lamellar stacking structures. Moreover, the surface topography of the N region displays some cracking flat tablets densely packed together, similar to the turtle shell. The morphology and cross-sectional SEM images of the AMS region are shown in Fig. 2(c) and (d). The myostracums with some narrow silts and voids can be easily observed in the AMS region, and the thickness of AMS region is about 15  $\mu\text{m}$ . Fig. 2(e) exhibits the surface morphology of the FP region, where the highly ordered prisms in a relatively large area can be observed. The inset in Fig. 2(e) indicates that these fibrous prisms are densely stacked. The size of prisms is approximately 1.5  $\mu\text{m}$  at average. These unique structures are beneficial to achieve the well SERS signal, since they can provide much rich hot spots while decorated with the noble metal. The cross-sectional SEM images of the FP region in Fig. 2(f) shows these prisms are entirely distributed in an oblique direction towards the shell margin.

In order to make these natural structures perform as plasmonic platforms, AgNPs were deposited on the M.c. by the PVD method. The SEM images of the N, AMS and FP regions on the AgNPs-M.c. substrate are respectively presented in Fig. 3(a)–(c). As we can see from the insets in Fig. 3(a)–(c), AgNPs has the dense arrangement (narrow nanogap is around 3 nm), and the average size is approximately 20 nm on these natural structures, which was measured with the Nano measurer software. Especially, the AgNPs likewise exist between the prisms densely, which indicates that the well three-dimension (3D) plasmonic structures could have formed. The EDS spectrum in Fig. 3(d) collected from the FP region on the AgNPs-M.c. substrate further confirms that the silver has been successfully deposited on the substrate.

The R6G was chosen as the analyte molecule to evaluate the SERS performance of these three plasmonic structures on the AgNPs-M.c. substrate. When using excitation in 532 nm, we can obtain the SERRS signal of R6G due to the resonance Raman contribution in R6G SERS spectra [40]. The Fig. 3(e) presents the SERRS signals of R6G molecules with the concentration of  $10^{-9}$  M from the three regions. From the typical spectrum collected from FP area on the AgNPs-M.c. substrate, the characteristic Raman peaks

of R6G at 610, 772, 1182, 1311, 1362, 1506 and 1647  $\text{cm}^{-1}$  could be observed clearly [40,41], and the intensity of each Raman bands is much higher than that in other two spectra collected from the AMS and N regions. The obvious bands at 1094 and 1483  $\text{cm}^{-1}$  all observed from three regions are assigned to symmetric stretching vibration mode and antisymmetric stretching vibration mode of  $\text{CO}_3^{2-}$  ions, respectively [29]. The intensity of Raman bands at 1094 and 1483  $\text{cm}^{-1}$  for three spectra have some differences, which is highly possible to be caused by the subtle difference for the composition of structures. As shown in Fig. 3(f), the peak intensity of 610 and 772  $\text{cm}^{-1}$  were quantitatively compared to estimate the SERRS effect of varied structures on the AgNPs-M.c. substrate. By calculation, the band intensity of 610  $\text{cm}^{-1}$  on FP region is approximately 17.5 times stronger than that on N region, 35.7 times higher than that on AMS region. Moreover, the band intensity of 772  $\text{cm}^{-1}$  on FP region are approximately 11.6 times stronger than that on N region, 30.6 times higher than that on AMS region. The high Raman enhancement of FP area on the AgNPs-M.c. substrate should be attributed to three major factors. First, AgNPs are densely decorated in the fibrous prisms (especially in the narrow gap between the adjacent prisms), forming 3D plasmonic structures which can provide much abundant hot spots. Second, the effective oscillate of incident laser can occur between the valleys of well-separated prisms, further enhancing the scattering cross-section. Third, the probing molecules could be likely to aggregate in the narrow space between the prisms. Besides, the cracks of NP region and the holes and hollows of AMS region on the AgNPs-M.c. substrate could also produce few hot spots, and the few probing molecule could slightly aggregate in these especial structures, so the low intensity of Raman bands can be still obtained. These results lead us to conclude that the FP region shows the better SERS performance than other regions on the AgNPs-M.c. substrate due to its high-ordered 3D plasmonic structures. Therefore, the substrates for the following measurements were all fabricated by the FP region of M.c.

To continue exploring the SERRS behavior of the FP region, we designed the GO-AgNPs-M.c. substrate. Fig. 4(a) and (b) represent the typical SEM images of the GO-AgNPs-M.c. substrate in



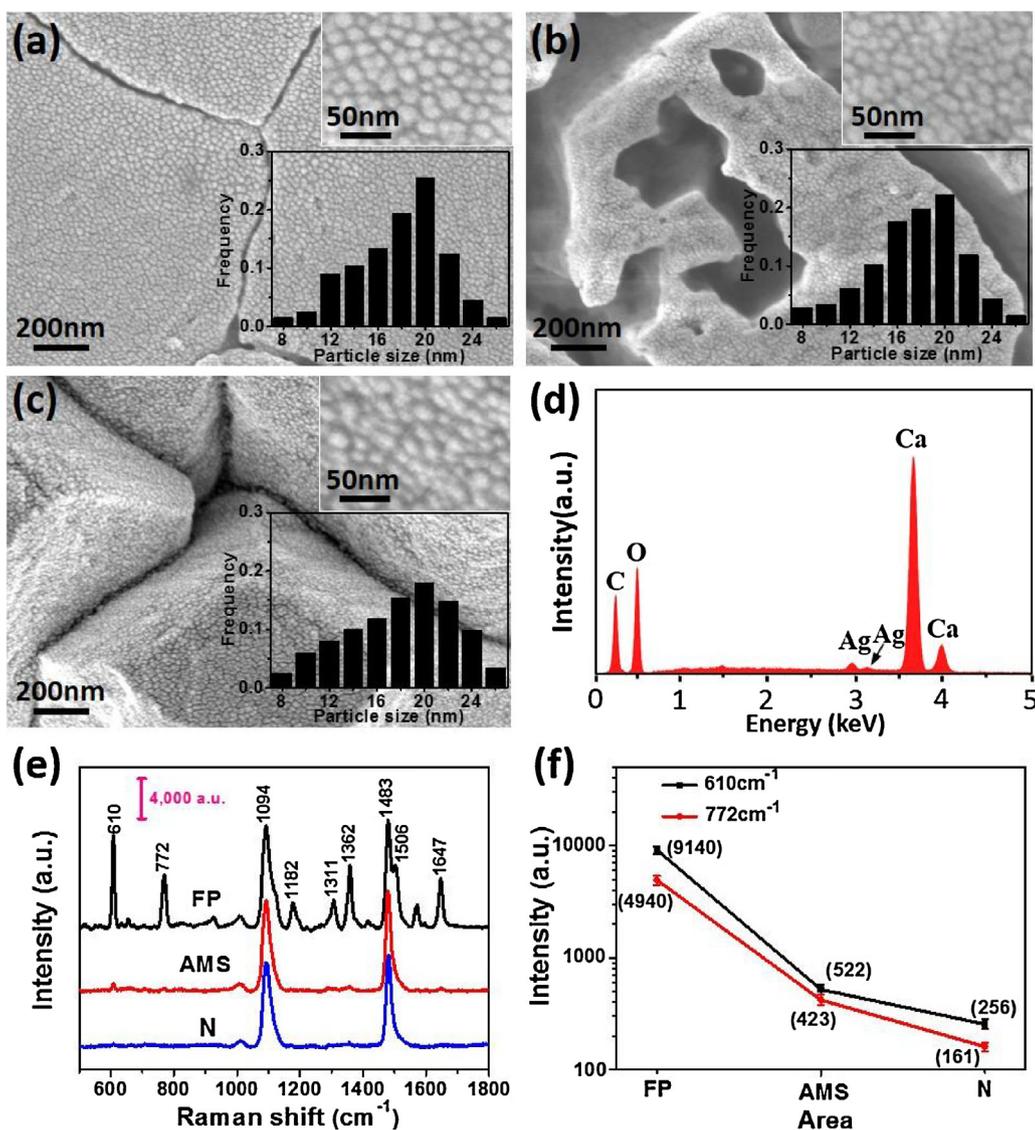
**Fig. 2.** (a), (c) and (e) represent the typical SEM images of the N, AMS and FP regions on the M.c., respectively. Insert in (e): SEM image of the FP region in a larger magnification. (b), (d) and (f) are the cross-sectional SEM images of the N, AMS and FP regions, respectively.

diverse magnifications. After the decoration of GO, the GO-AgNPs-M.c. substrate still displays typical pyramid-like fibrous prisms. The wrinkles of GO films could be obviously identified between prisms (marked with red arrows), proving that AgNPs-M.c. is covered by the GO film evenly. The origin Raman spectra of the AgNPs-M.c. and GO-AgNPs-M.c. substrates were further measured to confirm the GO decoration, as shown in Fig. 4(a). Apart from the original Raman peaks of AgNPs-M.c. substrate, the GO-AgNPs-M.c. substrate shows two obvious Raman peaks at  $1350$  and  $1587\text{ cm}^{-1}$ , which are related to the D and G bands of the disordered GO, respectively [21,39]. This result further reveals the successful modification of GO on the GO-AgNPs-M.c. substrate. In Fig. 4(d), the Raman mapping of  $1350\text{ cm}^{-1}$  peak of GO were obtained over an area of  $20 \times 20\ \mu\text{m}^2$  on the GO-AgNPs-M.c. substrate. The small color variations of the D peak demonstrated a relatively uniform distribution of GO in large scale.

Fig. 5(a) and (b) illustrate the SERRS signals of R6G with various concentrations on the AgNPs-M.c. and GO-AgNPs-M.c. substrates. All the characteristic vibrations of R6G can be confirmed with the previous mentioned, and the intensity of these bands decay with the decreasing concentrations monotonously. The enhancement effect of the GO-AgNPs-M.c. substrate is clearly superior to the AgNPs-M.c. substrate in term of the intensity of bands. Due to the

successful modification of GO, the small bulges of the Raman bands at  $1350$  and  $1585\text{ cm}^{-1}$  appear obvious on the GO-AgNPs-M.c. substrate. Importantly, the detection limit of the GO-AgNPs-M.c. substrate could be down to  $10^{-14}\text{ M}$ , which is one order of magnitude lower than that of the AgNPs-M.c. substrate. The better SERRS behavior of GO-AgNPs-M.c. hybrids can be ascribed to the following factors: first, the GO can provide the extra enhancement for the SERRS signal from the CM, which is correlated to the charge transfer between the substrate and the target molecules. Second, the GO film could reduce the loss of the target molecules to certain extent. The little partial target molecule solution could inevitably flow into the bottom of prisms along the small gaps between the prisms, inducing the reduction of the effective target molecules, further resulting in the unsatisfactory detections. Third, the AgNPs are closely covered by the GO, so the analyte molecules could be more effectively enriched around the hot spots, further resulting in more sensitive SERRS signals [39].

The relationships between the SERRS intensity of  $610\text{ cm}^{-1}$  peak and the concentration of R6G for the GO-AgNPs-M.c. and AgNPs-M.c. substrates are plotted in log scale in Fig. 5(c), where the linear fit curve can be observed. The values of  $R^2$  can reach 0.976 and 0.968 respectively, indicating that these two substrates could provide the quantitative analyze for the concentration of the analyte molecule.



**Fig. 3.** (a)–(c) are respectively the SEM images of the N, AMS and FP regions on the AgNPs-M.c. substrate. The insets in (a)–(c) are respectively the magnified SEM images and histograms of particle size distribution of the AgNPs. (d) The EDS spectra collected from the FP region on the AgNPs-M.c. substrate. (e) Measured SERRS signals of R6G with the concentration of  $10^{-9}$  M from three regions. (f) The relative SERRS intensity of the peaks at 610 and  $772\text{ cm}^{-1}$  in cases of three regions in log scale.

The former  $R^2$  is a bit higher than the latter  $R^2$ , attributing to the even distribution of R6G molecules with the existence of GO film on the GO-AgNPs-M.c. substrate.

Fig. 5(d) shows the SERRS signals of  $10^{-9}$  M R6G on the GO-AgNPs-M.c. and AgNPs-M.c. substrates, the resonance Raman signal of  $10^{-3}$  M R6G on blank  $\text{SiO}_2$  substrate separately. The enhancement distribution of GO was further quantified by the calculation of the enhancement factor (EF) according to the following formula [25–27,42,43]:

$$EF = \frac{I_{\text{SERS}}/N_{\text{SERS}}}{I_{\text{RS}}/N_{\text{RS}}} = \frac{I_{\text{SERS}}}{I_{\text{RS}}} \times \frac{N_{\text{RS}}}{N_{\text{SERS}}}$$

where  $I_{\text{SERS}}$  and  $I_{\text{RS}}$  represent the peak intensity of the SERS signal and the normal Raman signal, and  $N_{\text{SERS}}$  and  $N_{\text{RS}}$  are the number of molecules on the substrates within the laser spot. The band inten-

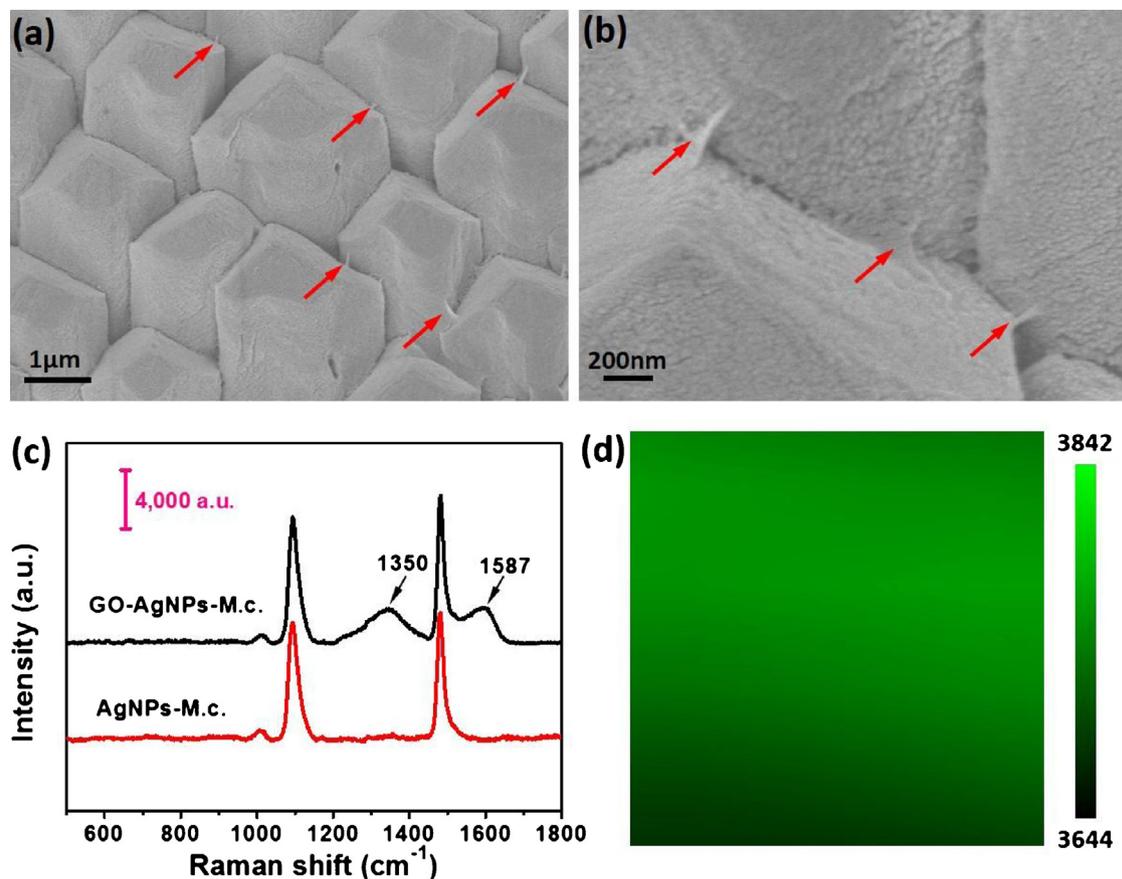
sity of  $610\text{ cm}^{-1}$  was chosen to calculate the EF. Therefore, the EF for the GO-AgNPs-M.c. hybrids is estimated:

$$\frac{16002\text{ (a.u.)}}{558\text{ (a.u.)}} \times \frac{10^{-3}\text{ (M)} \cdot 2\text{ (}\mu\text{L)} \cdot N_A \cdot \left(\frac{A_{\text{laser}}}{A_{\text{SiO}_2}^{\text{dry}}}\right)}{10^{-9}\text{ (M)} \cdot 2\text{ (}\mu\text{L)} \cdot N_A \cdot \left(\frac{A_{\text{laser}}}{A_{\text{SERS}}^{\text{dry}}}\right)} \approx 2.4 \times 10^7$$

where  $N_A$  is Avogadro's constant,  $A_{\text{SiO}_2}^{\text{dry}}$  is the dried area on the  $\text{SiO}_2$  substrate,  $A_{\text{SERS}}^{\text{dry}}$  is the dried area on the SERS substrate,  $A_{\text{laser}}$  is the area of laser spot. The dried area on  $\text{SiO}_2$  substrate is approximately 1.2-time larger than that on SERS substrate. Similarly, the EF for the AgNPs-M.c. hybrids is calculated:

$$\frac{7021\text{ (a.u.)}}{558\text{ (a.u.)}} \times \frac{10^{-3}\text{ (M)} \cdot 2\text{ (}\mu\text{L)} \cdot N_A \cdot \left(\frac{A_{\text{laser}}}{A_{\text{SiO}_2}^{\text{dry}}}\right)}{10^{-9}\text{ (M)} \cdot 2\text{ (}\mu\text{L)} \cdot N_A \cdot \left(\frac{A_{\text{laser}}}{A_{\text{SERS}}^{\text{dry}}}\right)} \approx 1.0 \times 10^7$$

Thence, the EF for the GO-AgNPs-M.c. substrate exhibits a 2.4-fold enhancement compared to the AgNPs-M.c. substrate, which agrees with the chemical enhancement caused by the GO layer [38].



**Fig. 4.** (a) and (b) are typical SEM images of the FP region on the GO-AgNPs-M.c. substrate in different magnifications. (c) Raman spectra on the GO-AgNPs-M.c. and AgNPs-M.c. substrates. (d) Raman mapping of  $1350\text{ cm}^{-1}$  peak of GO on the GO-AgNPs-M.c. substrate over an area of  $20 \times 20\ \mu\text{m}^2$ .

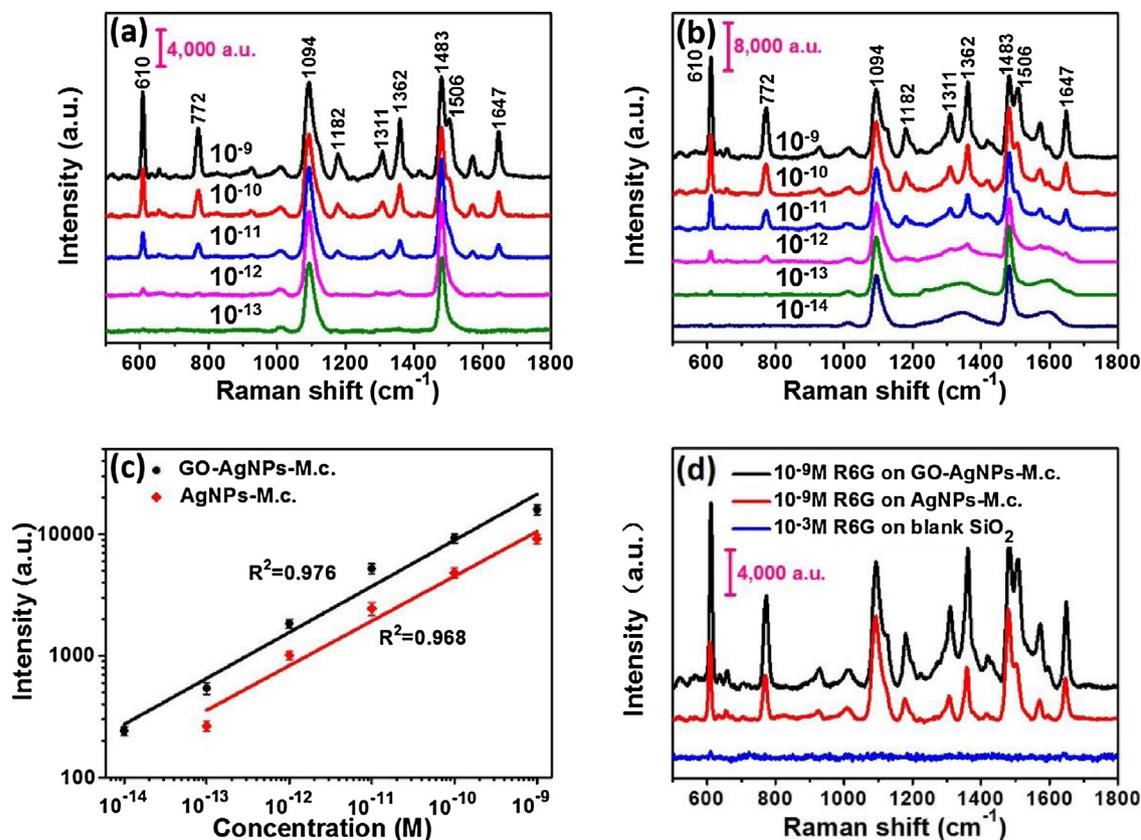
Considering that the GO mainly amplifies the Raman signal via the CM mechanism, and it has little effect on the electric field, so we utilized the COMSOL (RF module) Multiphysics software to simulate the electric field distribution on the AgNPs-M.c. structure. In the simulation, we set the particle size of AgNPs as 20 nm and the gap of the adjacent AgNPs as 3 nm. Fig. 6 shows the x-z view of the local electric field distribution on the AgNPs-M.c. structure that the adjacent prisms are arranged with angle at  $90^\circ$  and  $60^\circ$ , respectively. The relatively intense local electric field is distributed in the narrow space between the adjacent prisms. The electric field enhancement contributed in case of the narrow space with angle at  $90^\circ$  is 10.1-fold, which is slightly lower than the 11.9-fold electric field enhancement provided with the narrow space with angle at  $60^\circ$ . With the angle decreasing, the closer nanoparticles at the bottom of the gap could contribute to the relatively higher electric field enhancement. Thence, by the calculated result, we conclude that the unique natural structure decorated with the dense AgNPs could provide the well electric field enhancement contribution.

As an efficient SERS substrate, aside from the outstanding sensitivity, the high reproducibility is another vital parameter for practical detection. Hence, the SERS signals of the R6G molecules with the concentration of  $10^{-9}\text{ M}$  were detected to evaluate the reproducibility of the GO-AgNPs-M.c. substrate. Fig. 7(a) displays the SERS signals collected from 20 positions randomly within an area of  $50 \times 50\ \mu\text{m}^2$  on the same GO-AgNPs-M.c. substrate. The profiles of the collected spectra are very similar with the detected position changing. Neither a significant shift of the characteristic Raman bands nor the obvious change of the peak intensity could be observed, showing the well reproducible property of the GO-AgNPs-M.c. substrate. Moreover, as illustrated in Fig. 7(b), the intense SERS sensitivity at  $610\text{ cm}^{-1}$  was quantified and plotted

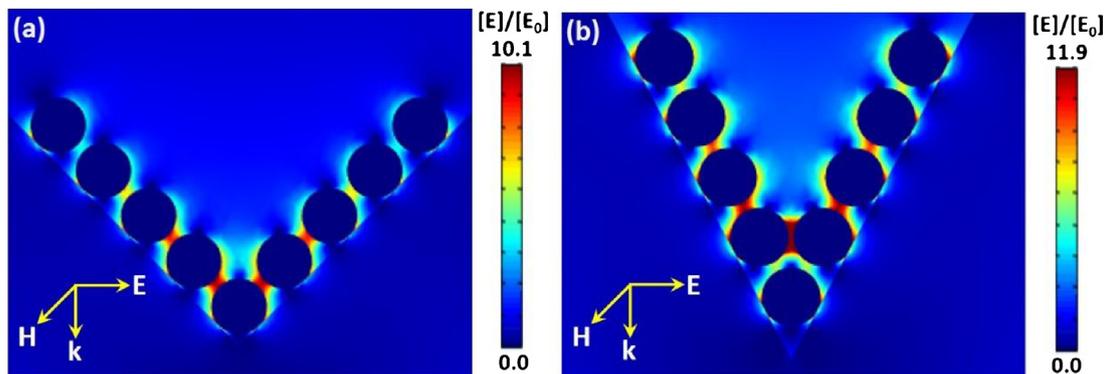
in the form of distribution histogram to analyze the relative standard deviation (RSD) [44]. The black horizontal line represents the average SERS intensity of  $610\text{ cm}^{-1}$  peaks from 20 positions, the shaded area shows the fluctuation of SERS signals. The RSD of the intensity of the  $610\text{ cm}^{-1}$  peak is 6.6% by calculation, further confirming that the GO-AgNPs-M.c. substrate possesses the good reproducibility. The results are attributed to the large-area well-ordered natural arrays and the uniform decoration of GO, making the GO-AgNPs-M.c. substrate act as the reproducible platform for the SERS applications.

For the fabricated GO-AgNPs-M.c. substrate, the GO was likewise introduced as a protective layer to prevent the oxidation of AgNPs. Therefore, the antioxidant stability of the GO-AgNPs-M.c. and AgNPs-M.c. substrates was investigated by the detection of R6G with the concentration of  $10^{-9}\text{ M}$ . Fig. 8(a) and (b) illustrate the SERS signals from two substrates under the exposure to the common air environment for 0 day and 15 days, separately. The peak intensity of  $610\text{ cm}^{-1}$  was chosen to compare the influence of the aerobic exposure for two substrates. By calculation, the decrease of the peak intensity on the GO-AgNPs-M.c. is only 8.4%. However, the peak intensity of  $610\text{ cm}^{-1}$  on the AgNPs-M.c. substrate has the drastic drop i.e. 44.7%, which is assigned to the decrease of the EM caused by the oxidation of AgNPs. In consequence, we can conclude that the GO decoration can endow the GO-AgNPs-M.c. substrate the well antioxidant stability on account of its superior chemical characteristic.

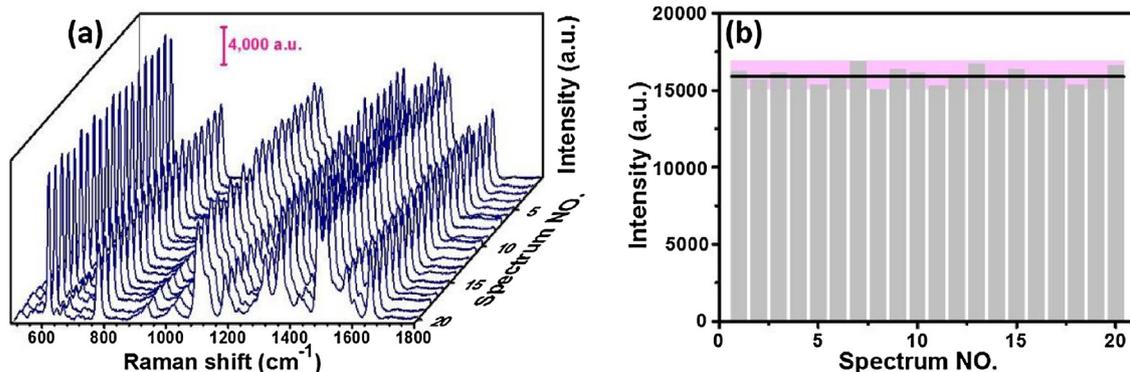
What's more, the feasibility of the GO-AgNPs-M.c. substrate in practical applications was further demonstrated by the detection of malachite green (MG) and methylene blue (MB). As a low-cost and highly efficient fungicide in the aquaculture samples, the high residual MG can cause severe harms to human body in terms of high



**Fig. 5.** (a) and (b) are respectively the SERS signals of R6G molecules with different concentrations on the AgNPs-M.c. and GO-AgNPs-M.c. substrates. (c) Raman intensity of 610  $\text{cm}^{-1}$  peaks as a function of different concentrations in log scale on the GO-AgNPs-M.c. and AgNPs-M.c. substrates. (d) SERS signals of  $10^{-9}$  M R6G on the GO-AgNPs-M.c. and AgNPs-M.c. substrates, the resonance Raman signal of  $10^{-3}$  M R6G on the  $\text{SiO}_2$  substrate.



**Fig. 6.** COMSOL-simulated x-z view of the local electric field distribution on the AgNPs-M.c. structure that the adjacent prisms are arranged with angle at (a) 90° and (b) 60°, respectively. E—electric field; H—magnetic field; k—direction of light propagation.



**Fig. 7.** (a) Collected SERS signals of R6G with the concentration of  $10^{-9}$  M from 20 randomly selected positions within an area of  $50 \times 50 \mu\text{m}^2$  on the GO-AgNPs-M.c. substrate. (b) The intensity variation of 610  $\text{cm}^{-1}$  peak for the GO-AgNPs-M.c. substrate.

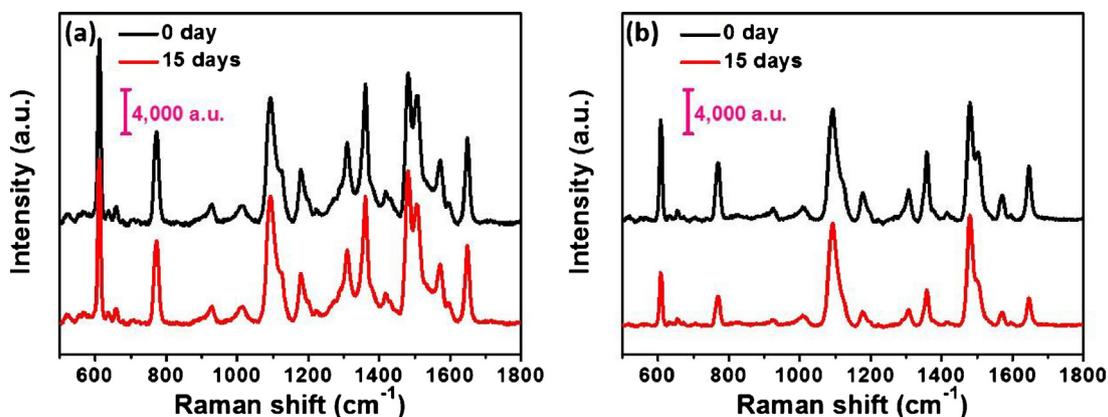


Fig. 8. (a) and (b) are respectively the SERS signals of R6G with the concentration of  $10^{-9}$  M on the GO-AgNPs-M.c. and AgNPs-M.c. substrates under the exposure to the common air environment for 0 day and 15 days.

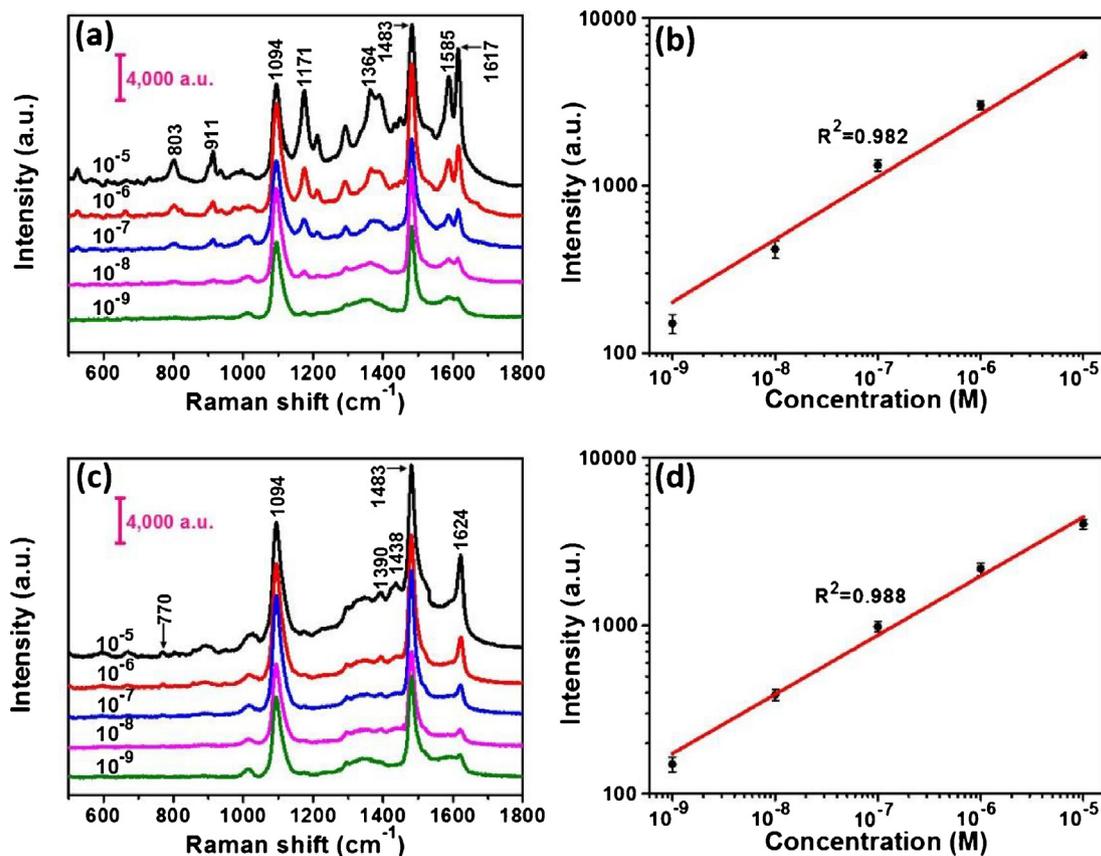


Fig. 9. (a) The SERS spectra of MG with different concentrations on the GO-AgNPs-M.c. substrate. (b) Raman intensity of MG at  $1617\text{ cm}^{-1}$  as a function of different concentrations in log scale. (c) The SERS spectra of MB with different concentrations on the GO-AgNPs-M.c. substrate. (d) Raman intensity of MB at  $1624\text{ cm}^{-1}$  as a function of different concentrations in log scale.

carcinogenicity, mutagenicity and teratogenicity [45,46]. Hence, the sensitive analysis of trace MG molecules plays a significant role in human health. The SERS spectra of MG with the concentrations range from  $10^{-5}$  to  $10^{-9}$  M were detected on the GO-AgNPs-M.c. substrate. Fig. 9(a) illustrates the dominating characteristic bands of MG in the region from 500 to  $1800\text{ cm}^{-1}$ . The primary Raman vibrations of MG are all observed in the spectra. The bands at 803, 911, 1171,  $1364\text{ cm}^{-1}$  are related to ring C–H out-of-plane bending, C–H out-of-plane bending and ring skeletal vibration, in-plane vibrations of ring C–H, and N-phenyl stretching vibrations respectively. The peaks located at 1585 and  $1617\text{ cm}^{-1}$  are assigned to ring C–C stretching vibration [47]. In addition, the related Raman

bands of the  $\text{CO}_3^{2-}$  ions and the GO could be still observed. The relatively strong SERS peak at  $1617\text{ cm}^{-1}$  was chosen to indicate the relationship between the presented SERS intensity and the MG concentration in log scale. The reasonable linear fit curve ( $R^2 = 0.982$ ) in Fig. 9(b) shows the excellent capability for the quantitative detection of MG. Besides, the sensitive detection of MB with the concentrations from  $10^{-5}$  to  $10^{-9}$  M further proves the well practicability of the GO-AgNPs-M.c. substrate. The MB has been reported as one highly toxic dye in various types of aquatic products. Adverse health effects caused by the MB toxicity include skin irritation, convulsions, tachycardia, dyspnea, cyanosis, renal failure, hyperbilirubinemia and hemolytic anemia [48,49]. As shown in Fig. 9(c),

the relatively intense characteristic peaks of MB were obtained in the region from 400 to 1800  $\text{cm}^{-1}$ . The relatively intense bands are in-plane bending mode of C–H at 770  $\text{cm}^{-1}$ , the symmetric and asymmetric C–N stretches at 1390 and 1438  $\text{cm}^{-1}$ , and the ring stretch at 1624  $\text{cm}^{-1}$  respectively [50]. As shown in Fig. 9(d), the  $R^2$  of the linear fit calibration curve for the peak at 1624  $\text{cm}^{-1}$  can reach 0.988, showing the well capability for the quantitative detection of MB. Therefore, the well sensitive detection of MB and MG show that the GO-AgNPs-M.c. hybrids possess immense potential for the practical applications in the environmental protection and food security.

#### 4. Conclusion

In summary, we fabricated an efficient natural SERS platform based on the GO-AgNPs-M.c. hybrids with a simply convenient and inexpensive method. With R6G as the analyte molecules, the FP region exhibits the most intriguing SERRS performance compared with other regions, which could be mainly attributed to abundant hot spots existed between mass uniform prisms. More importantly, the GO-AgNPs-M.c. hybrids can achieve highly sensitive SERRS measurements with brilliant reproducibility and antioxidant stability. It has a minimum detectable concentration for R6G as low as  $10^{-14}$  M. The well electric field contribution is confirmed in theory via the COMSOL software. Besides, the GO-AgNPs-M.c. hybrids still obtain the excellent reproducibility with a RSD of 6.6%. The experimental results demonstrate that the covered GO film not only provide an additional enhancement for the SERS signal from CM, but also ensure the stability of the GO-AgNPs-M.c. hybrids from the oxidation. In addition, this novel SERS natural substrate could be still successfully applied in the detection of MG and MB molecules. Therefore, the GO-AgNPs-M.c. substrate, as an effective SERS platform, can pave a new way to facilitate practical SERS applications in diverse biochemical fields.

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